Tropical Zoology

Geographical variation in the morphology and sexual dimorphism of Acanthochelys spixii (Testudines, Chelidae) in Brazil

A. Bager, P.S. Lucas, A. Costa, J.C.S. Lima & M.L. Silveira





Geographical variation in the morphology and sexual dimorphism of Acanthochelvs spixii (Testudines, Chelidae) in Brazil

Initial

Alex Bager^{*}, Priscila Silva Lucas, Aline Costa, Júlio Cézar Santos Lima and Melise Lucas Silveira

Departamento de Biologia, Setor de Ecologia, Universidade Federal de Lavras, Lavras, MG 37200 5 000, Brazil

(Received 02 June 2015; accepted 03 February 2016)

Body size is an important parameter for ecology, reproduction, evolution, and development of animal species, besides understanding their taxonomic relationships and to establishing the relationships between individual size and shape. We evaluated the variation in body size traits, sexual dimorphism in populations of a turtle species Acanthochelys spixii across its distribution in Brazil and we described sex ratio in those populations. Description of the size-classes on these populations was also provided. We sampled 86 adults (16 in Minas Gerais (MG) and 70 in Rio Grande do Sul (RS)). The sex ratio was 1:1 in both sampling areas. Females were larger than males, and specimens from RS were larger than those from MG. The posterior-lobe width (PLW) was deemed the most significant sexing variable in a linear discriminant analysis of specimens within each state. Two variables, PLW and maximum carapace width, together distinguished the four groups (males and females in MG and RS) with 77% accuracy. Overall, most body measures were larger for both males and females in the southernmost population, inhabiting higher latitudes and lower temperatures. Regarding sexual size dimorphism, females had larger PLW than males in MG, and all variables were dimorphic in RS, in general females being larger than males.

Keywords: Chelidae; sex ratio; morphometrics; sexual dimorphism; Acanthochelys spixii

Introduction

Body size is an important parameter that can affect the ecology, reproduction, evolution, AO1 and development of animal species (Preston and Ackerly 2004). Morphometric studies in turtles have contributed to understanding their taxonomic relationships and to establishing the relationships between individual size and shape, life history, and habitat characteristics (Daza and Páez, 2007). Populations of species that inhabit a wide range of environments frequently display divergent morphologies that correlate with differences in ecological pa-30 rameters (Rivera 2008).

In broader spatial scales, body size variation can be explained by environmental temperatures, known as Bergmann's rule (Mayr 1956). It is often cited as an example of adaptative geographic variation (Mayr 1956) in which there is a within-species tendency for increasing body size with increasing latitude or decreasing environmental temperature (Ashton and Feldman 2003). Other sources also could explain these variations, for instance, seasonality (Ashton et al. 2007). Seasonality is associated with body size, but not in a linear way so that smaller body sizes occur at the intermediate gradient of the predictor variable

© 2016 Istituto per lo Studio degli Ecosistemi of the Consiglio Nazionale delle Ricerche, Firenze.

25

10

15

20

^{*}Corresponding author. Email: abager@dbi.ufla.br

TTZO 1164559	Initial	CE: XX	QA: XX
17 March 2016	Initial	Coll:XX	QC:XX

(Ashton et al. 2007). Flow regimes (Rivera 2008) and anthropic effects such as hunting pressure, which can select for a given size class (Daza and Páez, 2007, Lovich et al. 2010), and interactions with other species, including predation, have also been examined.

40

Of interest, and also widespread described in many taxa (Shine 1979, Vollrath and Parker 1992, Owens and Hartley 1998, Cox et al. 2007, McPherson and Chenoweth 2012), sexual size dimorphism (SSD) is found in many reptile species (Anderson and Vitt 1990, Cox et al. 2003). These size differences, however, do not follow a general rule (Cox et al. 2007). In lizards and crocodilians, males are larger than females in most groups (Olson et al. 2002, Cox et al. 2007); for turtles and snakes the inverse seem to be the most general (Shine 1994, Cox et al. 2007). There is evidence that the observed patterns of reptile SSD for large male size are due to sexual selection and for large female size are due to fecundity selection (Shine 1994, Cox et al. 2003, 2007) and ecological divergence between sexes due to intraspecific competition (Preest 1994). Acanthochelys spixii (Duméril & Bibron, 1835), which belongs to the family Chelidae, is found in South American regions as Uruguay and Argentina (Carreira et al. 2007, Fritz and Havas 2007, Estrades et al. 2008). In Brazil its distribution ranges since central areas passing through southeastern to south region. (Brandão et al. 2002, Ribas and Monteiro Filho 2002, Fritz and Havas 2007, Brasil et al. 2011). The species inhabits slow-moving water bodies, ponds, wetlands, and other areas with abundant submerged vegetation (Quintela and Loebmann 2009). The biology and ecology of A. spixii are poorly understood, and most studies have been performed on specimens in captivity (D'amato, 1992, Brandão et al. 2002, Mocelin et al. 2008, Brasil et al. 2011). Among the publications describing the biology and ecology of A. spixii, none AQ2 has described the morphology of the species in detail. Biometric studies are an important tool for differentiating species and subspecies (Lamb and Lovich 1990, Lovich and Lamb 1995, Seidel et al. 1999), measuring geographical variation within species (Iverson 1985, Lubcke and Wilson 2007), analyzing reproductive parameters (Congdon and van Loben Sels 1991, Daza and Páez, 2007), and evaluating the impact of commercial exploitation (Close and Seigel 1997, Gamble and Simons 2004). Thus, a more detailed understanding of geographic variation in morphology is critical to elucidating the systematic relationships and ecological characteristics of this species.

In the present study, we sampled populations of *A. spixii* in two Brazilian states separated by nearly 2000 km in order answer the following questions: (i) are there differences in body size across populations? (ii) Do populations differ in sex ratio? and (iii) Do populations show sexual dimorphism?

Materials and methods

Study area

Acanthochelys spixii individuals were captured in two Brazilian states, Rio Grande do Sul (RS) and Minas Gerais (MG) (Figure 1). Sites were chosen based on previous species records and environmental peculiarity in each one of them. In RS, specimens were collected at six locations from 1994 to 2006, but not annually (Table 1). This region is highly homogeneous and typically of low elevation (mean = 4 m), with a geological origin in the Quaternary. The predominant aquatic habitats are puddles and ponds, notably the Lagoa Mirim and Lagoa dos Patos (Krause et al. 1982) (Figure 1). In MG, specimens were collected at Sempre-Vivas National Park (PARNA das Sempre-Vivas) in 2011. The sampling points were located in regions of the upper São Francisco River Basins, all at elevations above 800 m.

TTZO 1164559	Initial	CE: XX	QA: XX
17 March 2016	Initial	Coll:XX	QC:XX



Figure 1. Sampling areas in each state. In Minas Gerias (PARNA das Sempre-Vivas) municipality of Diamantina. In Rio Grande do Sul in the municipalities of Pelotas, Arroio Grande, Rio Grande and the Ecological Station Taim (ESEC Taim).

Data collection

Adult *A. spixii* specimens were captured using two types of bait trap: fyke-net traps (FN), with a single opening measuring 30.4 cm in width and 14.7 cm in height, and crab traps (CT), with four openings averaging 40.5 cm around the perimeter (Fagundes et al. 2010). Active searching in the water (ASW) and in the field (ASF) (Bager et al., 2007) and manual

- 5
- (CT), with four openings averaging 40.5 cm around the perimeter (Fagundes et al. 2010). Active searching in the water (ASW) and in the field (ASF) (Bager et al., 2007) and manual capture via snorkeling (SN) (Bager et al., 2010) were also demonstrated (Table 1), but here used only for purpose of morphometric description of the populations. Each individual was weighed (10-g accuracy), and up to 27 additional measurements were taken using a vernier caliper (1-mm or 0.1-mm precision). Figure 2 shows how the measurements were
- ¹⁰ made and their acronyms. Cephalic and mouth width were measured over the tympanic membranes and the maximum width of the lower rhamphotheca, respectively. Males were sexed based on secondary sexual characteristics: tail length (which is longer in males) and plastron concavity. The capture site of each individual was recorded using GPS, and all turtles were marked by notching the marginal scutes (Cagle 1939, Bager et al. 2010) and
- ¹⁵ then returned to their capture sites.

TTZO 1164559	Initial	CE: XX	QA: XX
17 March 2016	mitiai	Coll:XX	QC:XX

Table 1. Sampling areas with geographic localities, sampling periods, and specimen-capture techniques. FN: Fyke-net traps; SN: manual capture via snorkeling; CT: crab traps; ASF: active searching in the field; ASW: active searching in the water. All coordinates in datum WGS84.

State	Study area	Latitude	Longitude	Sampling period	Capture methods
MG	Diamantina (PARNA das Sempre-Vi- vas)	17°36′00″	44°60′00″	May 2010	FN/SN
RS	Taim Ecolog- ical Station (ESEC Taim)	32°32′51″	52°32'40"	1995, 1998, 2000, 2002, 2003	ASF/CT
	Pelotas	31°43′46″	52°21′51″	2002, 2003, 2004, 2006	ASW
	Arroio Grande	33°14′21″	53°08′12″	2001, 2002	ASW
	Rio Grande	32°06′00″	52°10′00″	1994, 1995	ASW

Data analysis

- ²⁰ All the comparative analyses were done based on individuals sampled by the traps and only measurements taken during the first capture were used in the analyses. Juveniles were disregarded due to the small number of individuals captured. Previous analysis did not identify morphometric variation between populations within RS state ($F_{2,47}$: 1.15, p = 0.28) and for this reason adults were grouped by sex: MG males, MG females, RS males, and
- ²⁵ RS females. Normality (Shapiro–Wilk test) and homogeneity of variance (Levene test) assumptions were checked. The Kruskal–Wallis test was used to compare different sexes within states (Zar 2010) (MG Male × MG Female; RS Male × RS Female) and the same sex from different states (MG Male × RS Male; MG Female × RS Female) in univariate analyses.
 ³⁰ The original non-transformed data ware used in the statistical analyses.

The original, non-transformed data were used in the statistical analyses, except in the discriminant analysis, in which the value of each variable was divided by the maximum length of the carapace. This approach was used to reduce the effect of size and to improve the allometric shape analyses between males and females (Bager et al. 2010).

Sexual dimorphism was analyzed within each region using linear regression (Zar 2010) with carapace length (CL) as the independent variable. Parallelism tests were used to compare the linear-regression results for males and females. When the parallelism test indicated a non-significant difference, we used covariance analyses (with CL as the covariate parameter) to compare the intercepts. In MG, the dependent variables were MCW, CCW, MPL, MVSL, MPW, PLW, MBL, CBL, CPD, CW, MW, and MHS. In RS, the dependent variables were MCW, MPL, MPW, PLW, CPD, and MHS.

The linear discriminant analyses (Zuur et al. 2007) of different sexes within states utilized seven variables (PLW, MPW, MHS, MVSL, MCW, MPL, and CPD) in MG and four variables (MCW, MPL, PLW, and MHS) in RS. A linear discriminant analysis was also used to compare the geographic variation in morphology among sexes and states, but only four variables (MCW, PLW, MPL, and MHS) were included in this analysis. The variables were selected to maximize the number of specimens in the analysis.

45

TTZO 1164559	Initial	CE: XX	QA: XX
17 March 2016	Initial	Coll:XX	QC:XX



Figure 2. Morphological measurements of *Acanthochelys spixii* (modified from Bager et al., 2010). Key: (1) carapace length (CL); (2) maximum carapace width (MCW); (3) central carapace width (CCW); (4) length of the third central scute (LC3); (5) width of the third central scute (WC3); (6) nuchal scute length (NL); (7) maximum carapace height (MHS); (8) carapace and plastron terminal distance (CPD); (9) maximum plastron length (MPL); (10) mid-ventral suture length (MVSL); (11) anterior lobe width (ALW); (12) maximum plastron width (MPW); (13) posterior lobe width (PLW); (14) central bridge length (CBL); (15) maximum bridge length (MBL); (16) width of left and right gular scutes (WGS); (17) left gular scute length (GSL); (18) left pectoral scute length (PEL); (19) left pectoral scute width (ABSW); (22) left anal scute length (ANSL); (23) left anal scute width (ASW); (24) mouth width (MW); (25) cephalic width (CW); (26) intergular scute length (ISL); (27) intergular scute width (ISW).

The sex ratio of males and females was analyzed by Chi-Squared tests (Zar 2010) using the Yates correction and equal expected proportions for both states. We used data from specimens captured only in ESEC Taim for RS state due to sampling method.





Figure 3. Frequency distributions of carapace length (mm) for male (dark bars) and female (gray bars) *Acanthochelys spixii* in Minas Gerais (A), and in Rio Grande do Sul (B).

SSD were quantified as recommended by Stephens and Wiens (2009):

SSD = (Mean Female CL / Mean Male CL) - 1

All analyzes were done using BioEstat 5.0 (Ayres et al. 2007). A significance level of $\alpha = 0.05$ was designated in all tests.

15 Results

In total, 85 individuals were collected: nine females and six males in MG and 38 females and 32 males in RS. Fifteen individuals were found at Sempre-Vivas National Park (Parque Nacional das Sempre-Vivas, in MG state) and the sex ratio (1.5:1) was not significantly different from 1:1 ($\chi^2 = 0.06$; p = 0.60). This parameter was also non-significant in ESEC Taim (RS state) (1.18:1, $\chi^2 = 0.51$; p = 0.47).

Male CL ranged from 101 to 178 mm, with a modal value of 150 mm, while female CL ranged from 134 to 180 mm, with a modal value of 160 mm (Figure 3). In general, females were larger than males in both regions (Table 2), but CL, MPW, and weight had similar size for females and males in RS, and the same was reported in MG state for the variables LC3, WC3, NCL, PEL, GSL, ISL, and ISW. The only variable for which males exceeded females in both the average value and the extreme values was ISW in MG; however, this difference was not significant.

$\begin{array}{ c c c c c c c c c c c c c c c c c c c$	RS							MG				RS/	MG
n Min $\bar{X} \pm 1SD$ CL 32 101-178 153.2 ± 17.3 MCW 32 80-125 108.3 ± 10.8 CCW - - - CCW 2 80-125 108.3 ± 10.8 CCW 32 80-125 108.3 ± 10.8 CCW - - - WC3 - - - WC3 - - - WC4 - - - WC5 - - - MC4 - - - MVSL - - - MVSL - - - MVSL - - - MPW 1100.1- 104.5 ± 4.8 MBL - - - MBL - - - MSL - - - MPW 1100.1- 104.5 ± 4.8 MBL		Female				Male			Femal	e		Male	Female
CL 32 101–178 153.2 ± 17.3 MCW 32 80–125 108.3 ± 10.8 CCW - - - LC3 - - - WC3 - - - MVL - - - MVSL - - - MVSL - - - MPU 1100.1- 104.5 ± 4.8 ALW - - - ALW - - - - MPU 100.1- 104.5 ± 4.8 - MLW - - - - MEU - - - -	u	Min.– Max	$\bar{X} \pm 1$ SD	P BS	и	Min.– Max	$\bar{X} \pm 1 \text{SD}$	и	Min.– Max	$\bar{X} \pm 1 \text{SD}$	P BS	Ρ	Р
MCW 32 80-125 108.3 ± 10.8 CCW - - - LC3 - - - LC3 - - - WC3 - - - WL - - - NL - - - MPL 32 96-168 141.2 ± 15.1 MVSL - - - MPU - - - MPU 100.1- 104.5 ± 4.8 MPW 7 100.1- 104.5 ± 4.8 MPW 7 112.66 - ALW - - - MPU 112.66 68.5 ± 8.2 PLW 31 45-83 68.5 ± 8.2 MBL - - - PLW - - - PSW - - -	35 1	34-176 1	159.0 ± 10.4	0.233	9	131-164	148.0 ± 10.6	6	152-180	163.9 ± 7.7	0.009	0.149	0.268
CCW - - - LC3 - - - WC3 - - - WL - - - NL - - - NL - - - MPL 32 96-168 141.2±15.1 MPW - - - MPW 7 100.1- 104.5±4.8 ALW - - - ALW 112.6 - - MPL 31 45–83 68.5±8.2 CBL - - - MBL - - - PSW - - -	35 1	01-132	114.8 ± 6.6	0.012	9	85-110	95.0 ± 8.4	6	103-116	107.3 ± 4.0	0.017	0.010	0.002
LC3	I	I	I	I	9	83-109	92.7 ± 8.7	6	94–115	102.6 ± 5.8	0.022	I	I
WC3	I	I	I	I	9	18.7–24.8	22.1 ± 2.8	6	22.5-27	24.5 ± 1.4	0.094	I	I
NL	I	I	I	Ι	9	23.5-29.7	25.5 ± 2.6	6	22.3-27.3	24.6 ± 1.5	0.738	I	I
MPL 32 96-168 141.2 ± 15.1 MVSL - - - MVSL - - - MVSL - - - MPL 100.1- 104.5 ± 4.8 MLW - - - ALW - - - PLW 31 45-83 68.5 ± 8.2 CBL - - - MBL - - - PEL - - - PSW - - -	1	I	18.3	Ι	9	11.6-14.9	13.7 ± 1.5	6	13.4-17.5	15.2 ± 1.3	0.111	I	I
MVSL - - - MPW 7 100.1- 104.5 ± 4.8 MLW - - - ALW - - - ALW - - - PLW 31 45-83 68.5 ± 8.2 CBL - - - MBL - - - PEL - - - PSW - - -	34 1.	28-165	150.4 ± 8.8	0.003	9	119-141	131.4 ± 8.3	6	138-164	148.3 ± 7.5	0.003	0.045	0.416
MPW 7 100.1- 104.5 ± 4.8 ALW - - - ALW - - - PLW 31 45-83 68.5 ± 8.2 CBL - - - MBL - - - PEL - - - PSW - - -	4 1	43-149	147 ± 2.8	I	9	113.9– 131.2	123.7 ± 6.5	6	129.6– 152.9	139.0 ± 6.4	0.003	I	0.034
ALW	6	91.2– 111.1	104.6 ± 5.8	0.427	9	I	I	I	I	I	I	I	I
PLW 31 45-83 68.5 ± 8.2 CBL - - - MBL - - - PEL - - - PSW - - -	4	39.7–93	90.5 ± 1.5	Ι	7	64.6-65	64.7 ± 0.3	1	Ι	73.7	Ι	Ι	Ι
CBL	34	63-89	77.2 ± 5.2	<0.001	9	57.5-69	62.0 ± 4.2	6	71-80.6	74.7 ± 3.1	0.001	0.021	0.064
MBL	I	I	I	I	9	25.7-31.3	28.3 ± 2.4	6	31.7-46.4	37.0 ± 4.4	0.001	I	Ι
PEL – – – – – – – – – – – – – – – – – – –	I	I	I	Ι	9	32.1–37.2	34.9 ± 2.2	6	37.1-49.7	43.0 ± 4.2	0.002	I	I
PSW – – – – – – – – – – – – – – – – – – –	4	11–19	13.4 ± 3.8	Ι	9	10.1–14.7	12.5 ± 1.6	6	9-16.6	13.1 ± 2.6	0.723	I	0.817
A CT	I	I	I	I	9	30.4–37.6	32.5 ± 2.7	6	37.4-41.2	39.3 ± 1.4	0.002	I	I
	I	I	I	I	9	17.3–21.8	19.8 ± 1.7	6	20.7-27.5	25.0 ± 2.0	0.004	I	I
ABSW – – – –	I	I	I	I	9	23.9–31.5	27.6 ± 2.5	6	30.8–38	34.8 ± 2.0	0.002	I	Ι
ANSL – – – –	I	I	I	I	9	18.2-22.5	20.2 ± 1.7	6	20-26.1	23.3 ± 2.0	0.015	I	I

Initial

TTZO 1164559

17 March 2016

(Continued)

CE: XX Coll:XX

QA: XX QC:XX

Table 2. ((Cont	'inued).														
				RS							MG				RS/I	МG
		Malo	e		Female	0			Male			Female	0		Male	Female
	и	Min.– Max.	$\bar{X} \pm 1$ SD	и	Min.– Max	$\bar{X} \pm 1 \text{SD}$	P BS	и	Min.– Max	$\bar{X} \pm 1$ SD	и	Min.– Max	$\bar{X} \pm 1 \text{SD}$	P BS	Р	Р
ASW	I.			1	1	1		6	21.9–25	22.7 ± 1.2	6	26.9-29.6	28.3 ± 0.8	0.001	I	I
GSL	T	I	Ι	I	I	I	Ι	9	17.1–19.3	18.3 ± 0.8	6	16-24.2	19.9 ± 2.8	0.221	I	I
WGS	T	I	I	T	I	I	I	9	50.1-58.8	54.6 ± 3.4	6	55.7-68.1	61.6 ± 4.1	0.005	Ι	I
ISL	Т	Ι	Ι	I	Ι	I	Ι	9	28-34.7	30.9 ± 2.5	6	29.3–37.4	32.3 ± 2.1	0.178	Ι	Ι
ISW	Т	I	Ι	I	I	I	Ι	9	17.2–21.3	19.0 ± 1.7	6	16.3-20.9	18.4 ± 1.8	0.358	I	I
CPD	\$	17.4–25.4	20.7 ± 3.5	6	24.8– 29.7	27.1 ± 2.1	0.009	9	16.1–23.3	21.3 ± 2.7	6	22.1–34.6	27.3 ± 3.5	0.004	0.927	0.775
CW	Т	I	Ι	б	28.3-30	29.4 ± 1.0	Ι	9	22.6–26	24.8 ± 1.3	6	26-29.6	27.5 ± 1.0	0.001	I	0.018
MW	T	I	Ι	I	I	Ι	Ι	9	17.4–21	19.1 ± 1.4	6	19.2-22.9	20.8 ± 1.3	0.030	I	I
SHM	30	42–68	54.4 ± 6.7	33	43-71	61.4 ± 6.8	<0.001	9	43-55	49.2 ± 4.3	6	52-60	55.6 ± 3.0	0.014	0.071	0.004
WEIGHT	ε	362-372	366.3 ± 5.1	e	540-668	595.3 ± 65.7	0.050	9	230-450	316.7 ± 74.5	6	340-530	433 ± 59.8	0.011	0.120	0.011

TTZO 1164559	Initial	CE: XX	QA: XX
17 March 2016	Initial	Coll:XX	QC:XX

The linear regressions for MCW, PLW, CPD, and MHS were significant and had high coefficients of determination, except for MHS in MG females ($F_{18} = 2.410$; p = 0.157; $r^2 = 0.14$) and CPD in RS males ($F_{14} = 0.443$; p = 0.555; $r^2 = 0.36$) (Table 3). Because this regression was not significant, analysis of covariance for CPD in RS should be regarded with caution. Overall, females had lower coefficients of determination than males, especially in RS.

The parallelism and covariance analyses to identify sexual dimorphisms in each state showed that PLW was the only dimorphic variable in MG ($F_{113} = 31.01; p < 0.001$), but all variables were dimorphic in RS. The number of individuals in MG certainly affected this result, and the analyses should be repeated with a larger number of specimens. Overall, SSD was 0.04 in RS and 0.1 in MG.

Linear discriminant analyses between sexes within states identified PLW as the only descriptor variable for sexual dimorphism in this species. The degree of explanation was 93% (Wilks' lambda = 0.3550; F_{113} = 23.617; p < 0.001) in MG and 78.7% (Wilks' lambda = 0.5804; $F_{1.59}$ = 42.660; p < 0.001) in RS.

15

5

10

The linear discriminant analysis comparing the four groups (sexes × areas) was highly significant (Wilks' lambda = 0.2640; $F_{6.144}$ = 22.709; p < 0.001) (Figure 4), and MCW and PLW were identified as the descriptor variables. The classification matrix had an overall

success of 77%. Females from MG were classified with 90% accuracy; the only misclas-20 sifications concerned RS females. Males from MG had the worst classification accuracy, 60%, but the small number of individuals (n = 6) affected this outcome. Females from RS were classified with 81% accuracy; they were sometimes confused with males from the same state and with females from MG. The model correctly classified 70% of RS males; the rest were confused with females from the same state.

25

Discussion

Despite its wide distribution in Brazil, A. spixii remains poorly studied. Data on its biology are found predominantly in the gray literature, and there have been several studies on its distribution (D'amato and Morato, 1991, Ghizoni et al. 2011), diet (Brasil et al. 2011), and demographics (Neto et al. 2011). Our results are consistent with previously published descriptions of the morphology and sexual dimorphism of A. spixii (D'amato and Morato, 1991, Bujes 2010, Neto et al. 2011). However, they are more comprehensive than previously available descriptions, enabling us to better assess the patterns of geographic variation.

Our data show that female A. spixii are significantly larger than males in most traits. Other authors have reported smaller measurements for A. spixii than those obtained here. Bujes (2010) reported a mean CL of 136.4 mm for females and 129.4 mm for males, and Neto et al. (2011) gave a mean size of 139.8 mm for females and 128.8 mm for males. The SSD values reported here are the first submitted for this species and demonstrate low dimorphism. The data reported by Neto et al. (2011) also indicate a low SSD (0.08), demonstrating that A. spixii follows the pattern of most Chelidae (excluding genera such as *Phrynops*) (Cox et al. 2007, Ceballos et al. 2012) in which females are larger than males. Smaller males could confer an advantage in agile courtship behaviors and mate searching in aquatic species in which male combat is absent, for example, (Berry and Shine 1980). In addition, females could grow larger than males because females gain an advantage in larger size to increase fecundity producing more or larger eggs or reproducing more frequently (Berry and Shine 1980, Stephens and Wiens 2009).

The sex ratios found in the present study confirm the results of Neto et al. (2011) and Bujes (2010), demonstrating a clear pattern of sexual structure throughout the species'

					Regressic	on parameters			Parall	lelism	Cov	ariance
	State	Sex	и	a	p	F	Р	24	F	Р	F	Р
MCW	MG	Male	9	-16.78	0.76	43.220	0.004	0.89	4.880	0.050	2.05	0.177
		Female	6	36.61	0.43	19.410	0.003	0.67				
	RS	Male	31	16.85	0.60	345.313	<0.001	0.92	3.124	0.079	10.11	0.003
		Female	34	41.34	0.46	37.696	<0.001	0.52				
PLW	MG	Male	9	13.86	0.33	9.280	0.039	0.62	0.099	0.757	31.01	<0.001
		Female	6	15.44	0.36	32.440	0.001	0.78				
	RS	Male	30	3.83	0.42	134.420	<0.001	0.82	0.305	0.590	39.38	<0.001
		Female	33	16.36	0.04	30.546	<0.001	0.47				
CPD	MG	Male	9	-10.43	0.21	10.600	0.032	0.66	0.490	0.506	2.87	0.116
		Female	6	13.42	0.08	0.290	0.606	-0.08				
	RS	Male	5	13.30	0.05	0.443	0.555	0.36	0.101	0.746	30.98	<0.001
		Female	8	2.51	0.16	12.087	0.010	0.58				
SHM	MG	Male	9	-5.55	0.37	20.440	0.012	0.79	1.401	0.263	1.16	0.306
		Female	6	24.81	0.19	2.410	0.157	0.14				
	RS	Male	29	4.66	0.33	65.722	<0.001	0.69	9.126	0.004	I	I
		Female	32	-32.06	0.59	56.428	< 0.001	0.63				

Initial

TTZO 1164559 17 March 2016

TTZO 1164559	Initial	CE: XX	QA: XX
17 March 2016	Initial	Coll:XX	QC:XX



Figure 4. Linear discriminant analysis between males and females of *Acanthochelys spixii* in the Minas Gerais and Rio Grande do Sul states. Triangle: Female MG; x: Female RS; Diamond: Male MG; Square: Male RS and Circle: Centroids.

range. In turtle species with genotypic sex determination, a 1:1 sex ratio is expected to occur (Janzen and Paukstis 1991) and this could be the case, as *A. spixii* is a Chelid species and most Chelid species follow this pattern (Ferreira 2009).

The two populations exhibited differences in size structure, especially for males. The modal class for this sex was 140 mm in MG and 150 mm in RS. No males smaller than 130 mm were captured in MG, preventing us from assessing whether secondary sexual dimorphisms (tail length and concavity) begin to develop at the same size found in RS (CL = 101 mm). Females had larger CPD measurements than males, in contrast to *Hy-dromedusa tectifera* (A. Bager, unpublished data), which shows higher CPD values for males (most likely resulting from the tail size). A possible explanation is that *A. spixii* is smaller in body size than *H. tectifera*, requiring a larger CPD size for spawning.

The linear regressions for CPD were not significant for MG females or RS males, while those for MHS were not significant for MG females, which had the lowest correlation coefficients among the analyses performed. This result is partly due to the small number of individuals sampled, but other measurements had higher coefficients despite an identical sample size. The parallelism and covariance analyses showed different results between the states. The variables MCW and PLW showed similar growth rates for both sexes in RS (with larger females) and different growth rates for the sexes in MHS. In MG, only PLW showed the larger size of females. This result was confirmed by the discriminant analysis, in which PLW was selected as a descriptor variable.

Comparisons within a given sex between study areas showed that both males and females were larger in RS in some related caparace and plastron measures, but not in others. Males in the two states were equal in size based on CL, CPD, MHS, and weight but differed in MCW and in plastron-related variables (MPL and PLW). Females in the two states were also similar in CL but showed no clear pattern in the variables that differed significantly between states, in contrast to the carapace, plastron, and head of males. Axis

TTZO 1164559 Initial 17 March 2016	CE: XX QA: XX Coll:XX QC:XX
------------------------------------	--------------------------------

I of the discriminant analysis separated the specimens by state, with MCW as the main group-descriptor variable. The second axis separated the specimens by sex, and PLW was the most significant variable. The discriminant analysis results confirmed that *A. spixii* is monomorphic, and width-related measurements were the descriptor variables.

5

The morphological differences between individuals may be linked to environmental factors, including air and water temperature and other habitat characteristics (Litzgus and Smith 2010), or to historical factors limiting gene flow between the basins (Clavijo-Baquet et al. 2010). However, the possibility that the morphological differences result from geographical isolation can be ruled out because the populations inhabit different basins instead of contiguous geographic areas (Clavijo-Baquet et al. 2010). Conversely, environmental

- 10 factors may have affected the morphological differences between the populations studied here because water and air temperature affect the activity and use of space by *A. spixii*. Furthermore, the behavior, breeding season, and feeding ecology of this species vary seasonally (Horta 2008), and the minimum air temperature, water temperature, and mean air temperature are the climatic parameters that best explain the variation in *A. spixii* move-
- 15 ment (Horta 2008). This study shows the geographic variation in which individuals from South populations (higher latitudes) are larger than the ones who inhabit lower latitudes for *A. spixii*. Further comparative studies should account for more populations to show how these morphological traits are related to temperature and other environmental conditions for this species in broader areas and quantitatively infer if in fact changes in shape and body size are directly related to environmental conditions.

Acknowledgements

We are grateful to Ramon Gomes de Carvalho to help us with some figures.

Disclosure statement

No potential conflict of interest was reported by the authors.

Funding

This work was supported by the Conselho Nacional de Desenvolvimento Científico e Tecnológico [grant number 303509/2012-0]; Fundação de Amparo à Pesquisa do Estado de Minas Gerais [grant number CRA – PPM-00139-14], [grant number CRA – APQ-03868-10].

References

- Anderson RA, Vitt LJ. 1990. Sexual selection versus alternative causes of sexual dimorphism in teiid lizards. Oecologia. 84:145–157.
- Ashton KG, Burke RL, Layne JN. 2007. Geographic variation in body and clutch size of gopher tortoises. Copeia. 2007:355–363.
- Ashton KG, Feldman CR. 2003. Bergmann's rule in nonavian reptiles: turtles follow it, lizards and snakes reverse it. Evolution. 57:1151–1163.

- Ayres M, Ayres M Jr, Ayres DL, Santos AS. 2007. Bioestat 50 aplicações estatísticas nas áreas das ciências biológicas e médicas. Belém: Sociedade Civil Mamirauá. 364 p.
- Bager A, de Freitas TRO, Krause L. 2007. Nesting ecology of a population of *Trachemys dorbignyi* (emydidae) in Southern Brazil. Herpetologica. 63:56–65.

AQ3

TTZO 1164559	Initial	CE: XX	QA: XX
17 March 2016	Initial	Coll:XX	QC:XX

- Bager A, Freitas TRO, Krause L. 2010. Morphological characterization of adults of Orbigny's slider *Trachemys dorbignyi* (Duméril & Bibron 1835) (Testudines, Emydidae) in southern Brazil. Tropical Zoology. 23:181–194.
- Berry JF, Shine R. 1980. Sexual size dimorphism and sexual selection in turtles (order testudines). Oecologia. 44:185–191.
- Brandão RA, Zerbini GJ, Sebben A, Molina FB. 2002. Notes on distribution and habitats of *Acan-thochelys spixii* and *Phrynops vanderhaegei* (Testudines, Chelidae) in Central Brazil. Boletín de la Asociación Herpetológica Española. 13:11–15.
- Brasil MA, de Freitas Horta G, Neto HJF, Barros TO, Colli GR. 2011. Feeding Ecology of Acanthochelys spixii (Testudines, Chelidae) in the Cerrado of Central Brazil. Chelonian Conservation and Biology. 10:91–101.
- Bujes CS. 2010. Os Testudines continentais do Rio Grande do Sul, Brasil:taxonomia, história natural e conservação. Iheringia. Série Zoologia. 100:413–424.
- Cagle F. 1939. A system of marking turtles for future identification. Copeia. 1939:170-173.
- Carreira S, Estrades A, Achaval F. 2007. Estado de conservación de la fauna de tortugas (Reptilia, Testudines) de Uruguay. Boletín Sociedad Zoológica Uruguay. 16:20–25.
- Ceballos CP, Adams DC, Iverson JB, Valenzuela N. 2012. Phylogenetic patterns of sexual size dimorphism in turtles and their implications for Rensch's rule. Evolutionary Biology. 2:194–208.
- 15 Clavijo-Baquet S, Loureiro M, Achaval F. 2010. Morphological variation in the South American snake-necked turtle *Hydromedusa tectifera* (Testudines: Chelidae). Chelonian Conservation and Biology. 9:231–237.
 - Close LM, Seigel RA. 1997. Differences in body size among populations of red-eared sliders (*Tra-chemys scripta elegans*) subjected to different levels of harvesting. Chelonian Conservation and Biology. 2:563–566.
- 20 Congdon JD, van Loben Sels RC. 1991. Growth and body size variation in Blanding's turtles (*Emydoidea blandingii*): relationships to reproduction. Canadian Journal of Zoology. 69:239– 245.
 - Cox RM, Butler MA, John-Adler HB. 2007. Sexual size dimorphism in reptiles. In: Fairbairn DJ, Blanckenhorn WU, Szekely T, editors. Sex, size, and gender roles: evolutionary studies of sexual size dimorphism. Oxford University Press; p. 38–49.

AQ5

- Cox RM, Skelly SL, John-Alder HB. 2003. A comparative test of adaptive hypotheses for sexual size dimorphism in lizards. Evolution. 57:1653–1669.
- D'amato ÂF. 1992. Notas sobre o desenvolvimento de *Platemys spixii* (Duméril e Bibron, 1835) (Testudines: Chelidae) em cativeiro. Acta Biologica Leopoldensia. 14:87–94.
 - D'amato AF, Morato SAA. 1991. Notas biológicas e localidades de registro de *Platemys spixii* (Duméril e Bibron, 1835) (Testudines, Chelidae) para o estado do Paraná. Acta Biológica Leopoldensia. 13:119–130.
 - Daza JM, Páez VP. 2007. Morphometric variation and its effect on reproductive potential in female Colombian slider turtles (*Trachemys callirostris callirostris*). Herpetologica. 63:125–134.
- ³⁰ Estrades A, Clavijo-Baquet S, Fallabrino A. 2008. Tortugas Dulceacuícolas del Uruguay. Almanaque. 2008:1–5.
 - Fagundes CK, Bager A. 2007. Ecologia reprodutiva de *Hydromedusa tecifera* (Testudines: Chelidae) no sul do Brasil. Biota Neotropica. 7:1–6.

AQ6

- Fagundes CK, Bager A, Cechín STZ. 2010. *Trachemys dorbigni* in an anthropic environment in Southern Brazil: I) sexual size dimorphism and population estimates. Herpetological Journal. 20:185–193.
- Ferreira PD Jr. 2009. Aspectos ecológicos da determinação sexual em tartarugas. Acta Amazonica. 39:139–154.
- Fritz U, Havas P. 2007. Checklist of chelonians of the world. Vertebrate Zoology. 57:149-368.
- Gamble T, Simons AM. 2004. Comparison of harvested and nonharvested painted turtle populations. Wildlife Society Bulletin. 32:1269–1277.
- Ghizoni IR Jr, Kunz TS, Cherem JJ, Bérnils RS. 2011. Registros notáveis de répteis de áreas abertas naturais do planalto e litoral do Estado de Santa Catarina, sul do Brasil. Biotemas. 22:129–141. Horta GF. 2008. Movimentação e uso do espaço por *Acanthochelys spixii* (Testudines, Chelidae) no

Parque Nacional de Brasília, Distrito Federal [master dissertation]. Brazil: Brasilia University. Iverson JB. 1985. Geographic variation in sexual dimorphism in the mud turtle *Kinosternon hirtipes*.

Copeia. 1985:388–393.

45

35

5

TTZO 1164559	Initial	CE: XX	QA: XX
17 March 2016		Coll:XX	QC:XX

Janzen FJ, Paukstis GL. 1991. Environmental sex determination in reptiles: ecology, evolution, and experimental design. The Quarterly Review of Biology. 66:149-179.

Krause L, Gomes N, Leyser KL. 1982. Observações Sobre a Nidificação e Desenvolvimento de Chrysemys dorbigni (Dumeril & Bibron, 1835) (Testudines, Emydinae) na Estação Ecológica do Taim, Rio Grande do Sul. Revista Brasileira Zoologia. 1:79-90.

Lamb T, Lovich JE. 1990. Morphometric validation of the striped mud turtle (Kinosternon baurii) in the Carolinas and Virginia. Copeia. 1990:613-618.

Litzgus JD, Smith SE. 2010. Geographic variation in sexual size dimorphism in Painted Turtles (Chrysemys picta). Journal of Herpetology. 44:320-326.

Lovich JE, Lamb T. 1995. Morphometric similarity between the turtles Kinosternon subrubrum hippocrepis and K. baurii. Journal of Herpetology. 29:621-624.

Lovich JE, Znari M, Baamrane MAA, Naimi M, Mostalih A. 2010. Biphasic geographic variation in sexual size dimorphism of turtle (Mauremys leprosa) populations along an environmental gradient in Moroco. Chelonian Conservation and Biology. 9:45-53.

Lubcke GM, Wilson DS, 2007, Variation in shell morphology of the western pond turtle (Actinemys marmorata Baird and Girard) from three aquatic habitats in northern California. Journal of Herpetology. 41:107-114.

Mayr E. 1956. Geographical character gradients and climatic adaptation. Evolution. 10:105–108.

McPherson FJ, Chenoweth PJ. 2012. Mammalian sexual dimorphism. Animal Reproduction Science. 131:109-122.

Mocelin MA, Fernandes R, Porto M, Fernandes DS. 2008. Reproductive biology and notes on natural history of the side-necked turtle Acanthochelys radiolata (Mikan, 1820) in captivity (Testudines: Chelidae). South American Journal of Herpetology. 3:223-228.

Neto HJF, Brasil MA, de Freitas Horta GF, Barros TO, Falcon GB, Colli GR. 2011. Demography of Acanthochelys spixii (Testudines, Chelidae) in the brazilian cerrado. Chelonian Conservation and Biology. 10:82-90.

Olsson M, Shine R, Wapstra E, Ujvari B, Madsen T. 2002. Sexual dimorphism in lizard body shape: the roles of sexual selection and fecundity selection. Evolution. 56:1538–1542.

Owens IPF, Hartley IR. 1998. Sexual dimorphism in birds: why are there so many different forms of dimorphism? Proceedings of the Royal Society B: Biological Sciences. 265:397-407.

Preest MR. 1994. Sexual Size dimorphism and feeding energetics in Anolis carolinensis: why do females take smaller prey than males?. Journal of Herpetology. 28:292–298.

Preston KA, Ackerly DD. 2004. The evolution of allometry in modular organisms. In: Pigliucci M, Preston KA, editors. Phenotypic integration: studying the ecology and evolution of complex phenotypes. Oxford (UK): Oxford University Press. p. 80-106.

Quintela FM, Loebmann D. 2009. Guia Ilustrado - Os répteis da região costeira do extremo sul do Brasil. Brazil: Manuais de Campo USEB.

Ribas ER, Monteiro Filho ELA. 2002. Distribuição e hábitat das tartarugas de água-doce (Testudines, Chelidae) do Estado do Paraná, Brasil. Biociências. 10:15-32.

10 Rivera G. 2008. Ecomorphological variation in shell shape of the freshwater turtle Pseudemys concinna inhabiting different aquatic flow regimes. Integrative and Comparative Biology. 48:769–787. Seidel ME, Stuart JN, Degenhardt WG. 1999. Variation and species status of slider turtles (Emydidae: Trachemys) in the southwestern United States and adjacent Mexico. Herpetologica. 55:470-487. Shine R. 1979. Sexual selection and sexual dimorphism in the Amphibia. Copeia. 1979:297–306.

Shine R. 1994. Sexual size dimorphism in snakes revisited. Copeia. 1994:326–346.

15 Stephens PR, Wiens JJ. 2009. Evolution of sexual size dimorphisms in emydid turtles: ecological dimorphism, Rensch's rule, and sympatric divergence. Evolution. 63:910–925. Vollrath F, Parker GA. 1992. Sexual dimorphism and distorted sex ratios in spiders. Nature. 360:

Zar JH. 2010. Biostatistical analysis. 5th ed. Upper Saddle River (NJ): Prentice-Hall. 960 p.

Zuur AF, Ieno EN, Smith GM. 2007. Analyzing ecological data. New York (NY): Springer. 672 p. 20

5

10

15

20

25

^{156-159.}